

THE PREVALENCE OF TRYPANOSOMA CRUZI, THE CAUSAL  
AGENT OF CHAGAS DISEASE, DETECTED IN RODENT  
HOST POPULATIONS IN TEXAS

by

Adriana Aleman

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Committee Members:

Dittmar Hahn, Chair

Michael R. J. Forstner

Ivan Castro Arellano

Hardin Rahe

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## **DEDICATION**

I dedicate this thesis to my parents, Tomas & Rosa Aleman, who provided me with their endless love, support and encouragement. Even though, they did not fully understand my project, they pushed me forward to reach my goals.

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## ABSTRACT

*Trypanosoma cruzi* is the parasite that causes Chagas disease, which affects over eight million people in at least 21 countries in Central and South America. While Chagas disease has been recognized as a significant health threat to the 28 million people living in Central America, it has not been historically considered a significant threat to the people in the United States. However, efforts to screen potential wildlife host populations for the parasite are only recently being undertaken in the southern USA. Since rodents are one of the reservoir hosts for *T. cruzi* and can be abundant close to human housing, detections of *T. cruzi* in rodents provide a good approximation of the prevalence of Chagas disease and the associated potential for the disease threat to human health. The purpose of this study was to determine the incidence of rodents infected with *T. cruzi* in five geographical regions across Texas, along with *Triatoma* vectors collected from three collection sites in Texas. DNA of the parasite *T. cruzi* was detected by real-time quantitative PCR (qPCR) in DNA extracted from heart tissue of rodents and the hindgut from *Triatoma* vectors, and prevalence assessed as a function of location, time of the season, and of rodent species. For the *Triatoma* vectors prevalence was assessed as a function of location, life stage and of *Triatoma* species. Of approximately 544 rodent samples analyzed, eight samples representing five rodent species were infected with *T. cruzi*. All of the positive detections of rodents occurred in the most southern geographical region of Texas, with significantly more detections in Winter compared to Spring and

Fall. Of thirty *Triatoma* vectors analyzed, 15 samples representing two *Triatoma* species were infected with *T. cruzi*. The data indicate that rodent and *Triatoma* populations in selected regions of Texas are infected with *T. cruzi*. Further studies need to be conducted to assess if other animal populations, or other rodent populations in Texas are infected with *T. cruzi* with the ultimate goal of understanding what the presence of this wildlife zoonotic means to human health in affected regions of our state.

# CHAPTER I

## INTRODUCTION

### *Background*

Chagas disease was discovered by the Brazilian doctor Carlos Ribeiro Justiniano Chagas in 1909 (WHO, 2014). However, even a century after its discovery, little is known about its distribution, prevalence and potential vectors in many countries.

*Trypanosoma cruzi* is the protozoa parasite that causes Chagas disease, which now affects over eight million people in at least 21 countries in Central and South America, including Mexico, Brazil, and others (CDC, 2013). Around 30-40% of the individuals infected with *T. cruzi* will develop symptoms of Chagas disease, which include cardiomyopathy (i.e. ventricular extrasystoles, ventricular tachycardia, and high degree heart block), digestive megasyndrome (i.e. dysphagia, odynophagia, and esophageal reflux) or both (Rassi, 2010; Bern, 2011). The remaining 60-70% of people infected will not develop specific symptoms, though signs of weakness, fever, inflammation and swelling of the site of inoculation can be prevalent (Bern, 2011). While Chagas disease has been recognized as a significant health threat to the 28 million people living in Central America, it has not been considered a threat to the people in the United States. In the United States, only about 300,000 people are estimated to have this disease (CDC, 2013). However, the disease is entering the U.S. from endemic countries through the migration of infected individuals (WHO, 2014). Other routes of distribution include the migration of animal reservoirs or vectors from countries having a high prevalence of animals infected with *T. cruzi* into countries without or having few documented

infections. In order to better understand this migration process both the wildlife reservoirs (e.g. rodent populations) and the vectors themselves (i.e. Triatominae bugs) provide an obvious source of data on the prevalence of the parasite in areas where human cases are unknown or very rarely reported.

Triatomine insects (Hemiptera: Reduviidae), commonly known as kissing bugs, are the vectors of transmission for *T. cruzi*. They are in the family Reduviidae, which contains 22 subfamilies (Kirchhoff, 1993). Triatomines are mainly found near woodpiles close to houses that are uninhabited (Coutinho, 2012). There are over 130 triatomine species that can carry *T. cruzi* in the Americas (Lent, 1979). There are about 11 species of triatomines in the United States, and seven of them have been found in Texas (CDC, 2013). These seven species are *Triatoma gerstaeckeri*, *T. indictiva*, *T. lecticularia*, *T. neotomae*, *T. protracta*, *T. rubida*, and *T. sanguisuga* (Kjos, 2009). There are five nymphal stages of triatomas from hatchling to adults (Galindez, 1998). Triatomines must intake a blood meal to develop through their nymph stages and into their adult stage (Bern, 2011). A blood meal is also required for the female triatomines to lay their eggs (Bern, 2011). The infection of the insect starts by the ingestion of a blood meal from an infected host. In the midgut of the insect, the *T. cruzi* parasites, trypomastigotes, transform into epimastigotes (Bern 2011). Trypomastigotes are the infective flagellated form of the parasite found in the blood of the mammalian hosts, and epimastigotes are the multiplying stage of the parasite that grows in the gut of the insect vector (Kirchhoff, 1993). Once in the hindgut, the epimastigotes then convert to metacyclic (i.e. infective) trypomastigotes, which are excreted in the feces. Once the infected insect takes another blood meal, they will then defecate near the bite wound. The metacyclic trypomastigotes

of *T. cruzi* can be transmitted from vector feces into an open wound site by the host scratching at the bite (de Freitas, 2011). Other ways that the disease can be transmitted are through blood transfusions, organ donations, and through congenital transmission (mother to child) (Grant, 1989).

There are two phases of Chagas disease, i.e. acute and chronic phases. The acute phase reflects the time from the initial bite and transmission to about 2 months after the initial infection. During this phase, individuals can be either asymptomatic or in some cases, have fever, headache, muscle pain and many other symptoms. The symptoms of an acute infection of Chagas disease can often be confused with a cold. If the infection is not treated while still in the acute phase, then it can become a chronic infection. It has been established that *T. cruzi* can enter and persist in the heart tissue of the host (Zang, 1990). During the chronic phase, individuals can suffer cardiac and digestive disorders. Some of the cardiac disorders are heart rhythm abnormalities, and a dilated heart, which can cause blood not to pump accordingly (CDC, 2013). This can lead to heart failure and thus death (Ferreira, 2011). The digestive disorders that can be seen are a dilated esophagus or colon, which can lead to problems eating and defecating (CDC, 2013).

There are no vaccines available to prevent Chagas disease (CDC, 2013). Treatment is only successful in patients with the acute form of the disease. Two medications can help with treating the infection, i.e. nifurtimox and benznidazole (Carod-Artal, 2013). These two medications, however, have very serious side effects (CDC, 2013). Moreover, they have not been approved by the Federal Drug Administration (FDA) in the United States, but can be obtained from the CDC under investigational protocols (CDC, 2013; Bern, 2011).

*Trypanosoma cruzi* species are genetically and biologically diverse (Campbell, 2014; Westenberger, 2005). Through multilocus genotyping techniques, *T. cruzi* revealed six discrete typing units (DTU) (Zingales, 2009). Molecular characterization separated *T. cruzi* into two main lineages, i.e. TcI and TcII (Anonymous, 1999; Zingales, 2009), and they seem to be recognized as the ancestral lineages (Westenberger, 2005). The *T. cruzi* II lineage can be further separated into five distinct subgroups TcIIa-TcIIe (Souto, 1996; Brisse, 2000; Brisse, 2001), which are also known as *T. cruzi* II to VI under recent nomenclature changes (Zingales, 2009). Due to these recent changes in the nomenclature, TcIIb became TcII, TcIIc became TcIII, TcIIa became TcIV, TcIId became TcV, and TcIIe became TcVI (Zingales, 2009). TcI and TcII are recognized as the ancestral lineages, whereas TcV and TcVI are hybrid lineages (Zingales 2009; Westenberger, 2005; Westenberger, 2006; de Freitas, 2006).

Diversity of parasite lineages among United States cases is not fully understood. In the United States there have been six *T. cruzi* autochthonous human cases with the first documented case of a female child in 1955 up to the most recent of an elderly female in 2006 (Woody, 1955; Dorn, 2007). Chagas disease was also discovered in a mummy over 1,150 years along the Rio Grande region in Texas (Reinhard, 2003). Of the six known genotypes of *T. cruzi* only two have been detected in the cases from the United States, i.e. TcI and TcIV, which have been identified from humans, wildlife, and vectors (Barnabe, 2001; Roelling, 2008). According to Roelling (2008), *T. cruzi* infected humans carry the genotype TcI, whereas *T. cruzi* infected wildlife and vectors contain the TcIV genotype.

The potential host diversity for the parasite is very broad as *T. cruzi* has been detected in over 100 mammalian species (Bern, 2011). All mammals are vulnerable to

this disease and can become reservoirs of *T. cruzi*. Animal reservoirs are mainly wild, free-ranging animals that are constantly exposed to *T. cruzi* vectors and that help to maintain this pathogen at a given locality. The first identified case of an infected wild animal in the US was a woodrat, *Neotoma macrotis*, in California (Charles, 2013). Today, there are reports of 24 species of wildlife, including armadillos, opossums, rodents, and woodrats, that have been found infected with *T. cruzi* in the U.S. (Charles, 2013). Many woodrat dens harbor triatomine insects. In some cases, other rodent taxa move into abandoned woodrat dens enabling infection and thus a broad array of potential reservoirs (Charles, 2013). Since rodents have been previously found infected with *T. cruzi* and can be abundant close to human housing, rodents will be the main focal point in this project. Rodents are also easily collected, can be obtained in large numbers, and thus detections of *T. cruzi* in rodents may enable a good approximation of the prevalence of Chagas disease. The results collected from this project will help to assess the prevalence of *T. cruzi* and thus the potential for Chagas disease in Texas.

There are many approaches to detect *T. cruzi* in animal reservoirs including serological, (i.e. enzyme-linked immunosorbent assay (ELISA), immunofluorescent-antibody (IFA)), and molecular tests (i.e. quantitative PCR) (Bern, 2011). In this project, a molecular tool was used to detect *T. cruzi*. This tool is quantitative real-time PCR (*qPCR*), a PCR-based method, which allows us to amplify and quantify a 166 bp fragment of satellite DNA of *T. cruzi* (Bern, 2011). Compared to serological tests, *qPCR* can detect *T. cruzi*, but also provides an accurate enumeration of *T. cruzi* (Bern, 2011). All positive samples obtained by *qPCR* analyses were re-amplified by PCR, the resulting

PCR products cloned and sequenced, and the sequences identified by comparative sequence analyses with those in established databases (i.e. Genbank).

### *Objective*

The basic goal of this research was to assess the prevalence of *T. cruzi* in rodent populations in Texas. Since the nests of rodents are often infested with triatomine bugs (Bern, 2011), they do not only provide reservoirs for *T. cruzi*, but are potential health hazards for humans. The prevalence of *T. cruzi* in woodrats has increased from the late 1900's to today (Charles, 2013). Thus, our basic hypothesis is that *T. cruzi* and thus a potential for Chagas disease is present in Texas, but more prevalent in southern areas of Texas closer to environments with naturally higher abundance of kissing bugs such as more tropical regions near Mexico. Due to lower abundance of kissing bugs and less favorable environmental conditions further north, it is believed that rodent populations in some northern regions in Texas do not harbor *T. cruzi*.

In order to test the hypothesis, rodents trapped in five geographical locations across Texas over a period of one year were tested for *T. cruzi*. The results were then assessed based on the detections as a function of location, seasonality, and rodent species. In this study, *Triatoma* vectors were tested for *T. cruzi* by collection of kissing bugs from three different locations in central Texas and the results were examined as a function of location, *Triatoma* species, and life stage.

## CHAPTER II

### MATERIALS & METHODS

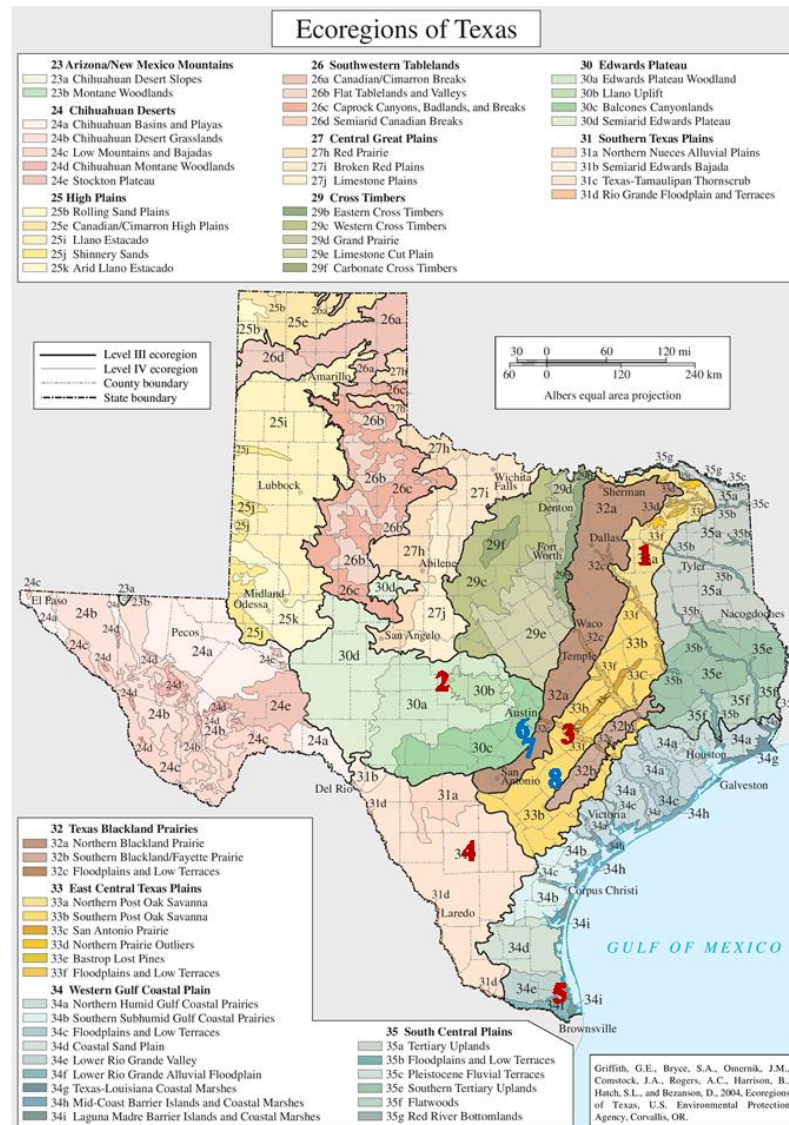
#### *Rodent Tissue Collection*

Rodents were collected from five sites covering 5 ecoregions in Texas as part of an ongoing project meant to quantify seroprevalence of zoonotic pathogens at rodent assemblages from contrasting habitats (Maikis, 2014; Milholland, in progress). All collection and handling of rodents was performed by Matt Milholland, Troy Maikis, and Dr. Ivan Castro-Arellano as enabled by Texas State University IACUC permit 1206\_0113\_02. The collection sites included one privately owned property, Tejas ranch (TR), and four Wildlife Management Areas (WMAs): Chaparral (CH WMA), Gus Engeling (GE WMA), Las Palomas (LP WMA), and Mason Mountain (MM WMA), which are managed by Texas Parks and Wildlife (Figure 1) (described in detail in Maikis, 2014). Each collection site was visited three times throughout the year and samples were collected in sylvan and disturbed habitats. Sylvan habitats displayed natural conditions of the property, while disturbed habitat displayed changes made by humans. The description of each site in both disturbed and sylvan areas are presented in Table 1, along with their designated ecological habitat. The first sampling period was in late winter, which started on February 9 and ended on April 7, 2013. The second sampling period was in spring, which started on April 19 and ended on June 14, 2013. The third sampling period was in the fall, which started on September 8, 2013 and ended on January 10, 2014. There were also a preliminary trapping period from January 25 to 26, 2013 in Mason Mountain WMA. The rodents were collected using 150 large folding aluminum Sherman live traps (H.B. Sherman Traps, Inc, Tallahassee, FL, USA), set up by sunset and checked and

closed after sunrise. The traps were set in linear arrangement six meters apart, and each contained a feed mixture of rolled oats, peanut butter, and imitation vanilla extract to attract rodents (Maikis, 2014).

#### *Triatoma Insect Collection*

*Triatoma* insects were collected from three different sites. In one site from San Marcos, TX (Edward Plateau, Hays County), the *Triatoma* insects were collected from wood piles in two week intervals from February to April and from September to November 2013. The second site was also in San Marcos, TX (Texas Blackland Prairie, Hays County), with two insects collected from inside a home in the month of June 2014. The third site was a collection from Gonzales, TX (East Central Texas Plains-Floodplains and Low Terraces, Gonzales County). All Gonzales county samples were collected at one time from a *Neotoma* den in October 2014.



**Figure 1:** Map of sampling sites. Rodent collection sites are: 1) Gus Engeling WMA (GE WMA), 2) Mason Mountain WMA (MM WMA), 3) Tejas Ranch (TR), 4) Chaparral WMA (CH WMA), and 5) Las Palomas WMA (LP WMA). *Triatoma* collection sites are: 6) San Marcos site 1, 7) San Marcos site 2, and 8) Gonzales.

**Table 1:** Description of field sites (Maikis 2014, Griffith, 2004).

Site	Ecological habitat	Disturbed	Sylvan
Gus Engeling Wildlife Management Area	East Central Texas Plains- Post Oak Savannah	Bunkhouse, learning center, maintenance yard, farm field	Natural Post Oak Savannah, prescribed fire recovery
Mason Mountain Wildlife Management Area	Edwards Plateau	Bunkhouses, decommissioned ungulate holding pens, maintenance facilities, man-made lake	Woodlands with heavy grassland
Tejas Ranch	East Central Texas Plains- Floodplains and Low Terraces	Farmhouse, barn, man-made lake, horse barn, deer feeder	Dry drainages, upland of forested floodplains, low terraces
Chaparral Wildlife Management Area	Southern Texas Plains	Unburned areas: main office, parking lot, burned areas: Bunkhouses, maintenance yard, outdoor storage	Areas untouched by the fire, and fire recovery areas
Las Palomas Wildlife Management Area	Western Gulf Coastal Plains	Original homestead, maintenance/storage yard, crop field, roadways	Dense, natural western gulf coastal plains vegetation

### *Sampling*

Rodents were collected from the traps and euthanized within 72 hours for the collection of external parasites as part of a previous study (Maikis, 2014). After anesthesia using cotton saturated with isoflurane, animals were euthanized by cervical dislocation. Total lengths, tail, ear, and hind foot length was measured, and gender and species of the rodent determined. Blood and organs (spleen, liver, heart, kidney, and knee joint) were collected from the rodent carcasses. Tissues were flash frozen using liquid nitrogen and maintained at -80°C to preserve tissue (Maikis, 2014). Heart tissue was made available for the study from this collection.

The insects collected in Hays County were frozen at -20°C, while those collected in Gonzales County were kept in ethanol at room temperature.

### *Analysis*

Heart tissue was used to determine the presence of *Trypanosoma cruzi* in the rodents. DNA from heart tissues was extracted using the DNeasy Blood and Tissue Kit (QIAGEN, Hilden, Germany) under Biosafety Level- 2 (BSL-2) conditions. A small sample of the heart, not greater than 25 milligrams, was used for DNA extraction. The samples were eluted with 400 µl of Buffer AE provided with the kit.

The intestinal content of the *Triatoma* insects were used to determine the presence of *Trypanosoma cruzi*. DNA was extracted using the DNeasy Blood and Tissue Kit (QIAGEN, Hilden, Germany) using BSL-2 conditions. A small sample of intestinal content, not greater than 25 milligrams, was used for DNA extractions. The samples less than 1.5 cm was eluted with 100 µl of Buffer AE, and samples greater than 1.5 cm was eluted with 200 µl of Buffer AE.

Following DNA extraction from rodents' heart and *Triatoma* gut, all samples were analyzed using quantitative PCR (*qPCR*) targeting a 166 base pair (bp) fragment of satellite DNA of *T. cruzi* (Bern, 2011). The standards that were used for the *qPCR* were PCR products obtained from a pure culture of *T. cruzi* (1987 or 2001) obtained from Universidad Autonoma del Estado de Morelos, Mexico. The 100 µl PCR reaction included: 62 µl of DI H<sub>2</sub>O, 16.5 µl of BSA (Thermo Fisher Scientific, Waltham, MA), 15mg/ml, 10 µl of 10x *Taq* Buffer and 0.2 µl final concentration of *Cruzi*-1 forward and *Cruzi*-2 reverse (300nM, Piron, 2007), 0.2 mM final concentration of dNTPs and 0.2 µl *Taq* Polymerase (GeneScript, Piscataway, NJ, US). The PCR conditions were as follows:

96°C for 10 minutes, and 45 cycles of 95°C for 15 seconds, and 58°C for 1 minute.

Samples were then loaded on a 2% agarose gel and the bands cut out and weighed. The bands were purified using the Ultra Clean 15 DNA Purification Kit (MO BIO Laboratories, Carlsbad, CA). PCR products were quantified using Qubit Fluorometric Quantitation (Thermo Scientific, Waltham, MA), and numbers of copies calculated (<http://cels.uri.edu/gsc/cndna.html>). Ten-fold dilutions starting with copy numbers of  $10^6$  to  $10^0$  were used as standards for detection and quantification.

The 10 µl *q*PCR mixture contained the following: 5 µl SYBR Green, 0.2 µl of 10 µM *Cruzi-1* forward, and 0.2 µl of 10 µM *Cruzi-2* reverse, 3.6 µl of H<sub>2</sub>O, and 1 µl of DNA solution. The *q*PCR conditions were 40 cycles at 95°C for 15 seconds and 58°C for 30 seconds. Samples from rodents and *Triatoma* insects were diluted tenfold and analyzed in a *q*PCR using the standards and the master mixture as stated. The dilutions were meant to evaluate whether there was any PCR inhibition from impurities in the samples. The samples were analyzed in triplicate. Using the DNA extraction of the *Triatoma* insects intestinal content, all *Triatoma* samples were analyzed using PCR (PCR) targeting mitochondrial COI. The 25 µl *Triatoma* PCR reaction included: 14.25 µl of DI H<sub>2</sub>O, 4.125 µl of BSA (Thermo Fisher Scientific, Waltham, MA), 15mg/ml, 2.5 µl of 10x *Taq* Buffer and 0.5 µl final concentration of forward and HCO 2198 reverse (Justi, 2014), 0.2 mM final concentration of dNTPs, 1.5 µl MgCl<sub>2</sub>, 3 mM, and 0.125 µl *Taq* Polymerase (GeneScript, Piscataway, NJ, US). The parameters for the thermal cycler were: 95°C for 5 minutes, and 34 cycles of 95°C for 30 seconds, 45°C for 45 seconds, 72°C for 1 minute: and 72°C for 10 minutes (Justi, 2014). Samples that did not provide results with a significantly high homology to sequences in the database were analyzed

using PCR targeting cytochrome B. The 25  $\mu$ l *Triatoma* PCR reaction included: 14.25  $\mu$ l of DI H<sub>2</sub>O, 4.125  $\mu$ l of BSA (Thermo Fisher Scientific, Waltham, MA), 15mg/ml, 2.5  $\mu$ l of 10x *Taq* Buffer and 0.5  $\mu$ l final concentration of 7432F forward and 7433R reverse primers (Pfeiler, 2006), 0.2 mM final concentration of dNTPs, 1.5  $\mu$ l MgCl<sub>2</sub>, 3 mM, and 0.125  $\mu$ l *Taq* Polymerase (GeneScript, Piscataway, NJ, US). The PCR conditions were the same as for the COI primers.

All positive samples were sequenced to verify the results obtained from *q*PCR. All samples from rodents and triatomas that were positive for *T. cruzi* were subjected to a PCR using the stated conditions above using the primers cruzi 1 and cruzi 2 targeting the 166 base pair segment of the satellite DNA (Piron, 2007). All *Triatoma* samples were analyzed in a PCR using the COI conditions as stated above. The samples that did not provide significant results were analyzed in a PCR reaction that targeted the Cytochrome B gene fragment (Pfeiler, 2006). PCR products were cleaned using Shrimp Alkaline Phosphate (Affymetrix, Santa Clara, CA, US) and Exonuclease I (Affymetrix, Santa Clara, CA, US) enzymes following the manufacturer's protocols. This was followed with bidirectional sequencing reactions using BigDye Terminator v3.1 (Applied Biosystems, Foster City, CA), with the same primers used for PCR. All samples were analyzed on a 3500 Genetic Analyzer for Resequencing and Fragment Analysis (Life Technologies, Carlsbad, CA, US).

Generalized linear mixed model was used to determine differences of function of location, or seasonality for the Texas samples.

## CHAPTER III

### RESULTS

#### *Rodents*

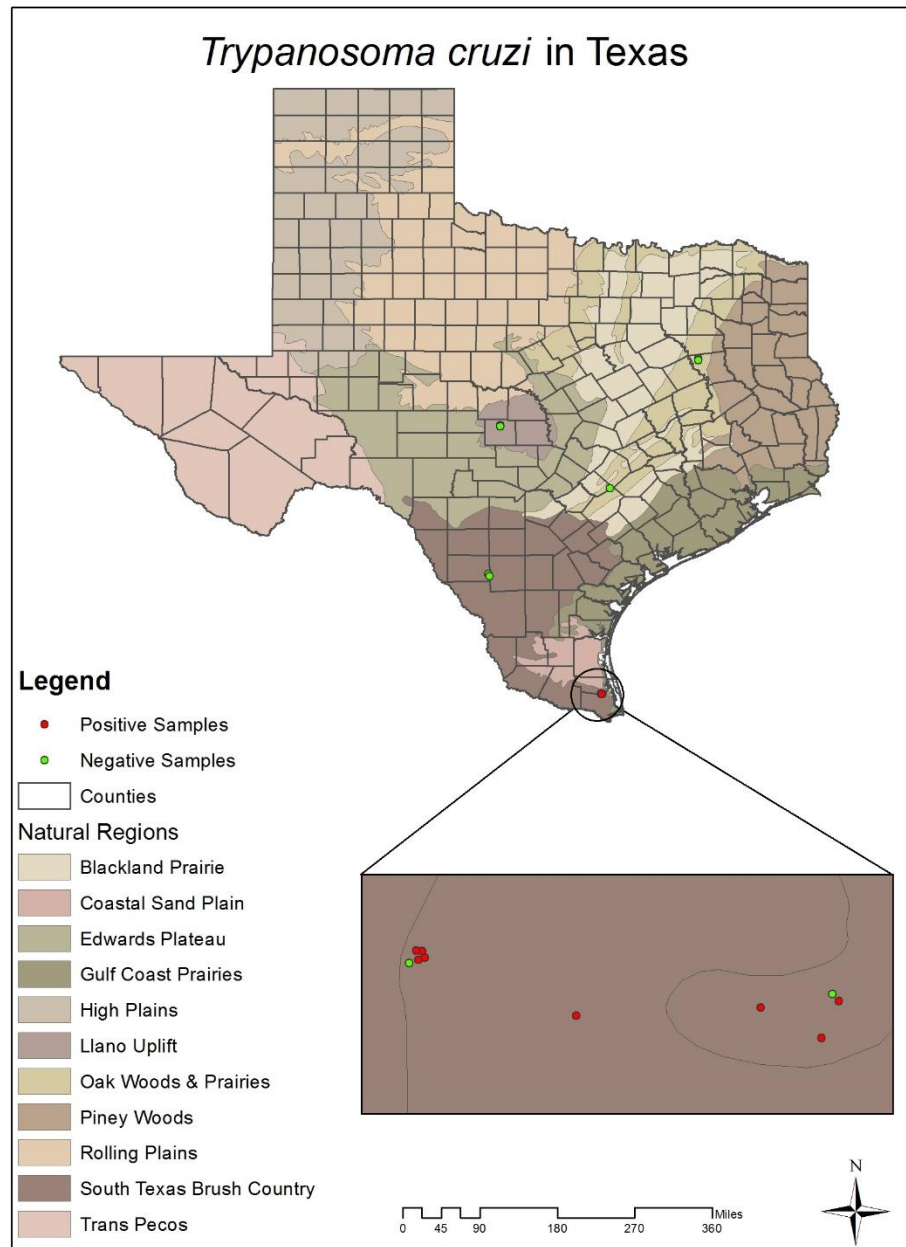
Eight out of 544 heart samples were infected with *T. cruzi* (Table 2). All eight infected rodents were from the same collection site of Las Palomas Wildlife Management Area, the most southern area sampled. Six positive samples were from the first collection in late winter (i.e. 16% of the Las Palomas samples). One positive was from the second collection in spring (i.e. 3% of the Las Palomas samples), and one from the last collection in winter (i.e. 1% of Las Palomas samples). Figure 2 depicts the locations where rodents were caught that were positive for *T. cruzi*. These rodents belonged to five species: *Neotoma micropus*, *Peromyscus leucopus*, *Sigmodon hispidus*, *Baiomys taylori*, *Liomys irroratus* (Figure 3). One rodent could not be identified to the species level, because *Peromyscus* sp. juveniles are particularly difficult to key from morphological characteristics. The disturbed habitat samples revealed four female rodents (4.71%) and one male (1.27%) positive for *T. cruzi* (Table 3). The sylvan samples revealed one female (.56%) and three males (1.69%) positive for *T. cruzi* (Table 3). Data indicate that differences were observed between seasons of harvest, with more positive samples observed in late winter compared to all other seasons ( $P= 0.05$ ). No significance was found when comparing gender within habitat.

**Table 2:** Comparison of frequency of rodents by *Trypanosoma cruzi* at five geographical location in disturbed and sylvan habitats.

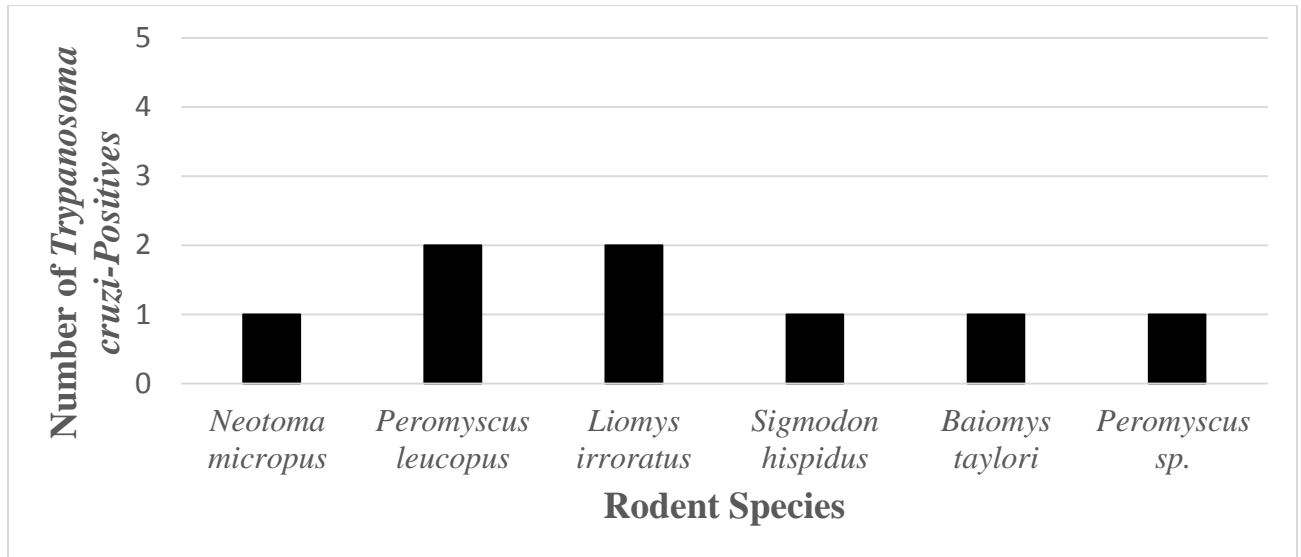
Site	Rodents collected <sup>1</sup>		Rodents positive for <i>T. cruzi</i>	
	Disturbed	Sylvan	Disturbed	Sylvan
Gus Engeling Wildlife Management Area	43 (13/28/2)	25 (12/8/5)	0	0
Mason Mountain Wildlife Management Area	49 (28/15/6)	52 (13/18/21)	0	0
Tejas Ranch	37 (12/9/16)	29 (9/13/7)	0	0
Chaparral Wildlife Management Area	94 (17/50/27)	24 (4/7/13)	0	0
Las Palomas Wildlife Management Area	129 (25/22/82)	37 (7/12/18)	5 (4/0/1)	3 (2/1/0)
Total	352	167	5	3

<sup>1</sup>numbers represent total numbers of animals and –in brackets- subsets collected in winter/spring/fall

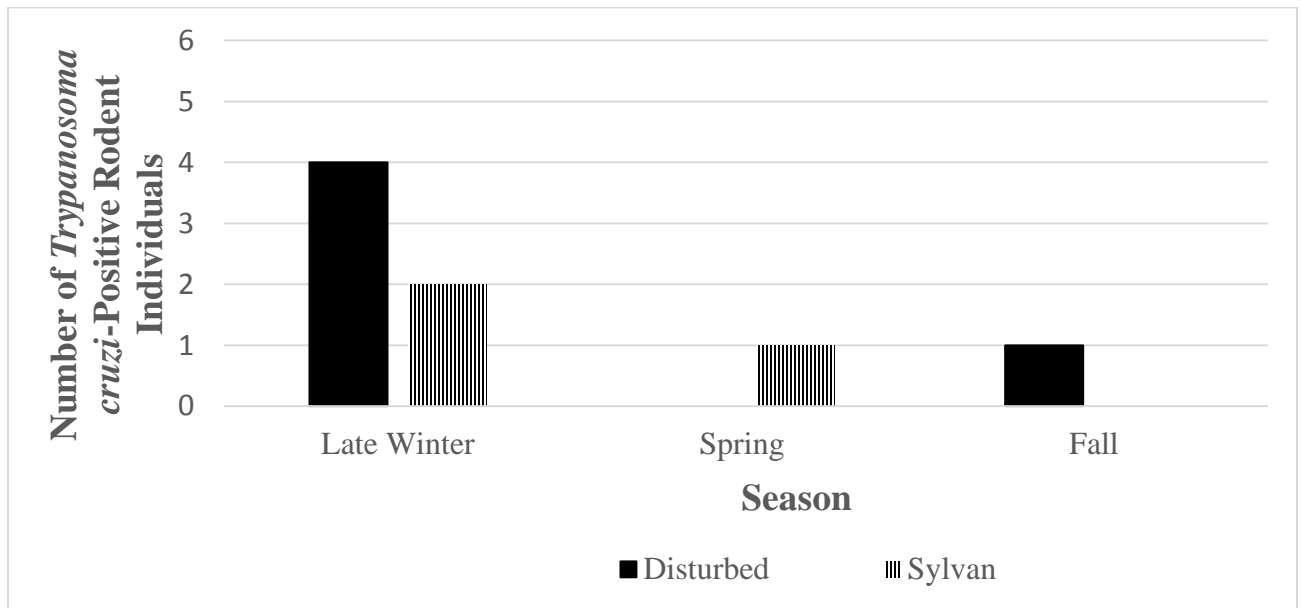
This table does not include preliminary samples collected from Mason Mountain.



**Figure 2.** Map showing positive samples at Las Palomas Wildlife Management Area. The green circles represent the negative samples. The red circles represents the positives in Las Palomas Wildlife Management Areas.



**Figure 3.** Number of *T. cruzi* infected rodents of *Neotoma micropus*, *Peromyscus leucopus*, *Liomys irroratus*, *Sigmodon hispidus*, *Baiomys taylori*, and *Peromyscus sp.*, which were detected using *qPCR*.



**Figure 4.** Comparison of the number of *Trypanosoma cruzi*-positive individuals between disturbed and sylvan habitats for three seasons sampled in Texas. There were more rodent's positives for *T. cruzi* in the late winter than spring and fall.

**Table 3:** Comparison of habitat by gender of rodents collected.

Habitat	Non-infected		<i>T. cruzi</i> Infected	
	Female	Male	Female	Male
Sylvan	93	84	1	2
Disturbed	179	180	4	1

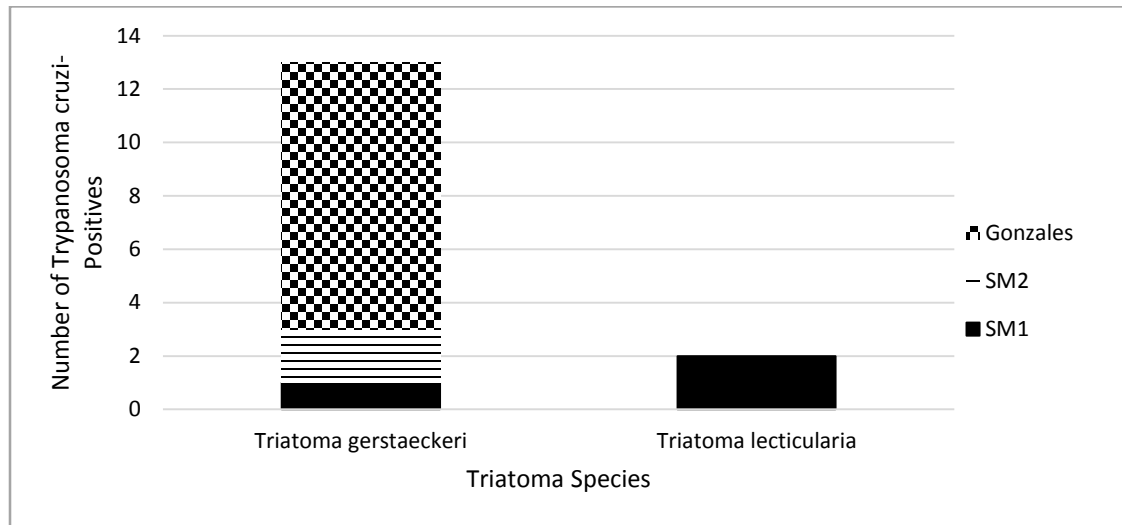
**Table 4:** Frequency of *Triatoma* vectors infected with *Trypanosoma cruzi* at three collection sites.

Site	<i>Triatoma</i> collected	<i>Triatoma</i> positive for <i>T. cruzi</i>
San Marcos Site 1	12	3
San Marcos Site 2	2	2
Gonzales	17	10
<b>Total</b>	30	15

### *Triatoma*

Fifteen out of 30 triatomine insects were shown to be infected with *T. cruzi*, as shown in Table 4. Three out of 12 *Triatoma* were positive from San Marcos site 1 (i.e. 25% San Marcos site 1). Two of two *Triatoma* were positive from San Marcos site 2 (i.e. 100% San Marcos site 2). Ten out of 16 *Triatoma* were infected with *T. cruzi* from Gonzales site (i.e. 62.5% Gonzales site). These *Triatoma* belong to two species: *Triatoma gerstaeckeri* and *Triatoma lecticularia*, as shown in Figure 5. The San Marcos site 1 contained three adults infected with *T. cruzi* out of eight adults and four nymph *Triatomas* (Table 5). These adults consisted of two *Triatoma lecticularia*, and one

*Triatoma gerstaeckeri*. The San Marcos site 2 contained two adult infected with *T. cruzi* out of two adult *Triatomas*, of the *Triatoma gerstaeckeri* (Table 5). The Gonzales site contained ten nymphs infected with *T. cruzi* out of 16 nymph *Triatomas* (Table 5). The positives obtained from this site consisted of the *Triatoma gerstaeckeri*.



**Figure 5:** *Triatoma* species positive for *Trypanosoma cruzi*.

**Table 5.** Comparison of sampling sites by life stage of *Triatoma* collected

Site	Non-infected		<i>T. cruzi</i> Infected	
	Nymph	Adult	Nymph	Adult
San Marcos site 1	4	5	0	3
San Marcos site 2	0	0	0	2
Gonzales site	6	0	10	0

#### *Triatoma* Species

Twenty-eight out of 30 *Triatoma* samples were analyzed using Cytochrome B and mitochondrial COI gene to determine their species. Two samples were unable to be amplified using either gene fragments. Nineteen *Triatoma* samples, three adults and 16 nymphs, matched *Triatoma gerstaeckeri* with a 99.4-99.8% pairwise identity. Two

*Triatoma* samples, two adults, matched *Triatoma lecticularia* with a 95.8% pairwise identity. Seven *Triatoma* samples, four adults and three nymphs, matched *Triatoma* sp. with a 99.1-99.9% pairwise identity.

#### *Trypanosoma cruzi* of Rodent Sequencing

PCR products were obtained from all positive rodent samples and directly sequenced. All PCR products were identical to each other and to the PCR product obtained from the two pure cultures, with three variable positions at the same positions. The variable positions were at bp positions 2, 87, and 116 of the 166 bp PCR product, and represented S(C/G), R(A/G) and K(G/T) substitutions, respectively. Genbank database searches of the consensus sequences revealed a 98.5% pairwise identity to *Trichomonas vaginalis* hypothetical protein (XM\_001294727) and a 98.4% pairwise identity to a *Trypanosoma cruzi* strain (HM01665).

Due to the three variability positions, PCR products from a rodent sample and two pure cultures were cloned into *E. coli*. Ten randomly selected clones from each sample were used as template for sequence analyses. Database searches for individual sequences showed up to 99.4% similarities of some of the clones to a *Trypanosoma cruzi* strain, but also to *Trichomonas vaginalis* hypothetical protein. The samples that matched to *Trichomonas vaginalis* still matched *Trypanosoma cruzi* with similar pairwise identities by a difference of 0.1%. These results confirm that our *qPCR* analyses specifically detect *T. cruzi*, and thus can be used to assess presence and abundance.

All clones matched to a *Trypanosoma cruzi* strain, but clones from one sample were quite diverse. Clones from the pure culture 2001 showed 100%-95% sequence similarity among each other, while those of pure culture 1987 revealed 100%-92%

sequence similarity values among each other. Clones from the positive rodent sample revealed 99%-94% sequence similarity values among each other. Comparative analyses of these sequences with those in the data base revealed 99-94% similarity values for pure culture 2001, 99-93% similarity for pure culture 1987 and 99-93% similarity for the positive sample with sequences of confirmed *Trypanosoma cruzi* strains. Clones from pure culture 1987 revealed 100-92% sequence similarities to clones from pure culture 2001, and those from pure culture 2001 100-93% sequence similarities to those from the positive rodent sample. Clones from pure culture 1987 revealed a 100-92% similarity to clones from the positive rodent sample. Thus, identical sequences were found between both pure cultures of *Trypanosoma cruzi* and the positive rodent sample, even though none of them had an identical sequence in the database.

#### *Trypanosoma cruzi* of *Triatoma* Sequencing

PCR products representing *T. cruzi* were obtained from all positive *Triatoma* samples and directly sequenced. Most sequences were similar, however, there were three samples from San Marcos site 1 to be different from all *Triatoma* samples. Genbank databases of consensus sequences revealed for the San Marcos site 1 a 98.7% pairwise identity to *Trypanosoma cruzi* strain (HM01665). Genbank databases of consensus sequence revealed for the San Marcos site 2 a 96.3-97.9% pairwise identity to *Trypanosoma cruzi* strain (EU178923). Genbank databases of consensus sequence revealed for the Gonzales site a 95.7-96.9% pairwise identity to *Trypanosoma cruzi* strain (EU178923).

## CHAPTER IV

### DISCUSSION

*Triatoma* sp. have been reported in many of the southern states within the United States (Bern, 2011). These states can potentially host individuals of *Triatoma* infected with *Trypanosoma cruzi*. *Trypanosoma cruzi* infected individuals have not been detected very often, but are becoming an increasing concern in the United States because the number of infected animals in Texas has been increasing. Thus, understanding the risks of the disease is increasingly important in Texas. Because it is difficult to detect and identify the disease in animal reservoirs, we focused on the detection of the causal agent, *T. cruzi*, as a proxy for the disease in this project. In this study, the only site that contained individuals infected with *Trypanosoma cruzi* was Las Palomas WMA. None of the sites in the northern, central, and eastern regions of Texas had any infected rodent host individuals among any of the 362 samples collected in the ecoregions. Many of the southern states, especially Texas, share neglected tropical diseases (NTD) with Mexico (Hotez, 2012), and Chagas disease, caused by *T. cruzi* is one of these diseases. This can be a function of similarity in climate, social environment, and the presence of *Triatoma* vectors that are common in the southern region of Texas and in Mexico (Hotez, 2012). While this study detected *T. cruzi* in a south Texas location, as seen in previous studies (Beard, 2003; Burkholde, 1980), it shows a slight northerly advancement from those detection sites. This can be a response to climate changes with increasing temperatures further north allowing many vertebrate hosts' and arthropod vectors moving north or toward higher elevations (Mills, 2010).

Rodents are one of the common hosts for *T. cruzi*, but very few studies have been done to view the prevalence of *T. cruzi* in the Texas rodent populations. *Trypanosoma cruzi* have been detected in 24 wild animal species within the United States so far (Zeledon, 2012). Within these 24 species, five were rodents captured in the state of Texas (Zeledon 2012; Burkholder, 1980; Barr, 1991; Kagan, 1966; Lathrop, 1965). In this current study, two rodent species that have been previously described as potential hosts of *T. cruzi*, i.e. *Neotoma micropus*, and *Liomys irroratus* (Burkholder, 1980) were found to host *T. cruzi* occasionally as well. We also found three additional rodent species that have not been previously described to carry *T. cruzi*: *Peromyscus leucopus*, *Sigmodon hispidus*, and *Biaomys taylori*. Several of these rodents are insectivorous, and might have their burrows infected with *Triatoma* vectors, thus increasing the potential exposure to *T. cruzi* (Eads, 1963). When wood rat habitats were analyzed for *Triatoma* vectors, a greater number of nymphs than adults were found inside the dens (Eads, 1963). This was also observed in this study at one of the sampling sites, which only contained nymphs with a high rate of infection with *T. cruzi*. This high concentration of nymphs in woodrat nests can be due to nymphs requiring blood meals to proceed into the next phase, until they reach adulthood, and thus having a source of blood easily available in dens benefits them because they are not required to find a new host. Rodents other than woodrats can become infected by moving into empty woodrat dens, which are likely to contain infected *Triatoma* (Bern, 2011).

In the Chaparral Wildlife Management Area, rodents' infected *T. cruzi* were not detected. However, in a previous study at the same collection site (Pinto 2010), 42 of 159 *Neotoma micropus* samples were found to be positive for *T. cruzi*. In our current study,

however, only seven *Neotoma micropus* samples were analyzed, which could explain the absence of any positive.

There were fifteen *T. cruzi* infected *Triatoma* out of 30 *Triatoma* samples collected. Of the fifteen positives, ten were nymphs and five adults. The ten nymphs came from the same collection site of Gonzales County. The other five positives were from two collection sites in Hays County. Both of these counties have had *T. cruzi* infected *Triatoma* (Kjos, 2009). Thirteen of the positives are from the *Triatoma gerstaeckeri*. Two positives from Hays County are *Triatoma lecticularia*. Three nymphs and four adults from Hays County are *Triatoma* sp.. The assignment of this *Triatoma* sp. was created by Kjos (2013) because comparative sequence analyses revealed high similarity to *Triatoma sanguisuga* and *Triatoma indictiva*, however, since no reference sequence for *Triatoma indictiva* was available in GenBank, they were unable to accurately identify the insects in question (Kjos, 2013).

By disturbing of the ecosystems in many sylvan habitats, humans can cause a higher likelihood to get in contact with the vectors (Sarkar, 2010; Mills 2010). *Triatoma* insects are attracted to homes that contain cracks in their foundation, walls, opening in their windows, and they are also attracted to strong light and carbon dioxide (Jurberg, 2006). These *Triatoma* insect are mainly active in the night, when they are taking blood meals from their host (Lent, 1979). Many humans are not aware of their presence, since they are only active at night, when most humans are asleep.

Comparative sequence analyses of cloned fragments of satellite DNA of *T. cruzi* from one of the PCR-positive samples and two pure cultures of *T. cruzi* revealed a highly variable fragment with multiple small sequence differences. Twenty-six clones were

sequenced for *T. cruzi* strain 2001 which resulted in 19 (73%) clones matching *T. cruzi* with a 97.5-99.4% pairwise identity, while the remaining seven (27%) clones matched a hypothetical protein in *T. vaginalis* with a 98.2-99.4% pairwise identity. Another 26 clones were sequenced for *T. cruzi* strain 1987 with 20 (77%) clones matching *T. cruzi* with a 95.6-99.5% pairwise identity, and six clones (23%) matching to *T. vaginalis* with a 98.2-99.4% pairwise identity. These clones also matched *T. cruzi* with the same identity. For a positive rodent sample obtained in this study, 40 clones were sequenced and 31 (77.5%) matched to *T. cruzi* while nine (22.5%) clones matched *T. vaginalis* with a 97-98.8% pairwise identity. One of these clones also matched *T. cruzi* with the same 98.2% pairwise identity. Although sequences of our two pure cultures and positive rodent samples did not match with 100% similarity to confirmed *T. cruzi* in the databases and some matched with high similarity values to a hypothetical protein from *T. vaginalis* leaves some questions on the identity (and thus the confirmation) of our detections of *T. cruzi*. However, individual clones of our pure cultures 2001 and 1987 matched to clones from our positive rodent samples 100%, and thus indicate the identity of our detections as *T. cruzi*. Additional studies, however, are needed that confirm these assumptions. These studies should include histological investigations as well as molecular analyses focusing on specific nucleic acid sequences other than the satellite DNA used in our investigations.

All three samples, i.e. the two pure cultures and the positive rodent sample, had at least one clone matching to a hypothetical protein of *T. vaginalis* (XM\_001294727). Hypothetical proteins are proteins predicted using computational methodology during genomic analysis (Sivashankari, 2006). Bioinformatics methods do not provide up to 70% of successful prediction accuracy (Bork, 2000). Thus, it is unlikely that the

prediction of the *T. vaginalis* gene is not accurate (Carlton, 2007). In the eukaryotic evolution, *Trichomonas vaginalis* branched earlier than trypanosomes, with the *T. vaginalis* containing hydrogenases instead of mitochondria like the trypanosomes (Housler, 1997). When comparing at *T. vaginalis* and *T. cruzi* targeting sequences, they share some sequence similarities (Housler, 1997). *T. vaginalis* and *T. cruzi* share some ancestral characteristics, and this might be the reason why in this study some clones matched sequences from *T. vaginalis*, even though more than seventy-five percent matched sequences from *T. cruzi*. *Trichomonas vaginalis* is protozoa parasite, which causes Trichomoniasis, a non-viral sexual transmitted disease (Van der Pol, 2007). *Trichomonas vaginalis* is mainly found in the genitourinary areas of humans (Harp, 2011). There have not been any cases or studies demonstrating the presence of *T. vaginalis* in rodent populations.

This project has detected a significant prevalence of infected rodent individuals at one site. These data indicate that *T. cruzi* infections are moving into higher latitudes which bears the attendant potential of *T. cruzi* and thus Chagas disease to become a significant zoonotic issue for Texas. As there are no vaccines or medication available to the public for prevention, the disease may prove to be difficult to control (Bern, 2011; Klotz 2014). The only path to preventing this NTD from becoming a major issue, is to educate individuals of the vector and the potential harm triatomines can create for domesticated animals and humans. This project was initiated to provide information on the prevalence of the parasite rather than focusing on the detection of the disease which is difficult to diagnose. By using two approaches, an assessment of the host reservoir in rodents and of the vectors themselves, I sought to evaluate the prevalence of *Trypanosma*

*cruzi* in Texas. Both methods provided positive evidence that the parasite is established in Texas and further research is needed to understand the distribution and impact from the parasite widespread presence in Texas. For future studies it would be important to look at the vectors at the same time as the host to enable comparisons of the parasite identity between them. Selecting more sampling sites across Texas, including regions in west Texas, would also enable a more comprehensive evaluation of the prevalence of *T. cruzi* in the state.

## APPENDIX SECTION

### Appendix I: *T. cruzi* infected Rodents and Control samples 166 base pairs fragment of satellite DNA

Consensus_Sequence	AST	CGG	CTG	ATC	GTT	TTC	GAG	CGG	CTG	CTG	CAC	CAC	ACG	TTG	TGG	TCT	ATG	TTT	TTG	TTT
TJM_163_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
TJM_185_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
TJM_188_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
TJM_192_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
TJM_196_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
TJM_435_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
TJM_628_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
1987_Positive_Control	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
2001_Positive_Control	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Consensus_Sequence	CGA	ATT	ATG	AAT	GGC	GGG	AGT	CAG	AGR	CAC	TCT	CTT	TCA	ATG	TAT	GTT	TGC	GTG	TKC	ACA
TJM_163_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
TJM_185_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
TJM_188_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
TJM_192_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
TJM_196_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
TJM_435_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
TJM_628_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
1987_Positive_Control	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
2001_Positive_Control	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...

Consensus_Sequence	CAC	TGG	ACA	CCA	AAC	AAC	CCT	GAA	CTA	TCC	GCT	GCT	TGG	AGG	AAT	T
TJM_163_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
TJM_185_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
TJM_188_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
TJM_192_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
TJM_196_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
TJM_435_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
TJM_628_PCR_Positive	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
1987_Positive_Control	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
2001_Positive_Control	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.

## Appendix II: *T. cruzi* infected *Triatoma* samples 166 base pairs fragment of satellite DNA

Consensus_Sequence	ACT	CGG	CTG	ATC	GTT	TTC	GAG	CGG	CTG	CTG	CAT	CAC	ACG	TTG	TGG	TCT	AGA	TTT	TTG	TTG
Gonzales_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_5	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_6	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_7	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	..C	...	...	...	...
Gonzales_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_11	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.A.	...	...	...
Gonzales_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_15	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
JAM_01	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Trial_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.A.	...	...	...
Trial_2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	..C	.A.	...	...	...
MF_5	...	...	...	...	...	...	...	...	...	...	..C	...	...	...	...	...	.TG	...	...	..T
MF_6	...	...	...	...	...	...	...	...	...	...	..C	...	...	...	...	...	.TG	...	...	..T
MF_13	...	...	...	...	...	...	...	...	...	...	..C	...	...	...	...	...	.TG	...	...	..T



### Appendix III: Cloned 166 bp satellite DNA fragments of pure cultures and rodent sample

Consensus_Sequence	ACT	CGG	CTG	ATC	GTT	TTC	GAG	CGG	CTG	CTG	CAC	CAC	ACG	TTG	TGG	TCT	ATG	TTT	TTG	TTT
Clone_2001.1.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.1.2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.3.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.3.2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.4.1	.G.	...	...	...	...	...	...	...	...	...	...	...	...	C..	...	...	...	...	...	...
Clone_2001.4.2	.G.	...	...	...	...	...	...	...	...	...	...	...	...	C..	...	...	...	...	...	...
Clone_2001.5.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.5.2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.6.1	.G.	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.6.2	.G.	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.8.1	.G.	...	...	...	...	...	...	...	...	...	...	...	...	...	..C	...	...	...	...	...
Clone_2001.8.2	.G.	...	...	...	...	...	...	...	...	...	...	...	...	...	..C	...	...	...	...	...
Clone_2001.9.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.9.2	.G.	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.10.1	.G.	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.10.2	.G.	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.11.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.11.2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.12.1	.G.	...	...	...	...	...	...	...	...	...	...	...	...	C..	...	...	...	...	...	...
Clone_2001.12.2	.G.	...	...	...	...	...	...	...	...	...	...	...	...	C..	...	...	...	...	...	...
Clone_2001.13.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.13.2	.G.	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_2001.14.1	...	...	...	...	...	...	...	T..	...	...	...	...	...	C..	...	...	...	...	...	...
Clone_2001.14.2	...	...	...	...	...	...	...	T..	...	...	...	...	...	C..	...	...	...	...	...	...
Clone_2001.15.1	...	...	...	...	...	...	...	...	...	...	...	...	...	C..	...	...	...	...	...	...
Clone_2001.15.2	...	...	...	...	...	...	...	...	...	...	...	...	...	C..	...	...	...	...	...	...

[illegible]

[illegible]

[illegible]

[illegible]

[illegible]

Consensus_Sequence	AGT	CGG	CTG	ATC	GTT	TTC	GAG	CGG	CTG	CTG	CAC	CAC	ACG	TTG	TGG	TCT	ATG	TTT	TTG	TTT
Clone_163.1.1	.C.	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.1.2	.C.	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.2.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.2.2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.3.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.3.2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.4.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.4.2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.6.1	...	...	...	...	...	...	...	...	...	...	...	...	...	C..	..C	...	.AA	...	...	...
Clone_163.6.2	...	...	...	...	...	...	...	...	...	...	...	...	...	C..	..C	...	.AA	...	...	...
Clone_163.7.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.7.2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.8.1	...	...	...	...	...	...	...	T..	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.8.2	...	...	...	...	...	...	...	T..	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.9.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.9.2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.10.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.10.2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.11.1	...	...	...	...	...	...	...	T..	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.11.2	...	...	...	...	...	...	...	T..	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.12.1	...	...	...	...	...	...	...	...	...	...	...	...	...	C..	...	.AC	.AA	...	...	...
Clone_163.12.2	...	...	...	...	...	...	...	...	...	...	...	...	...	C..	...	.AC	.AA	...	...	...
Clone_163.13.1	...	...	...	...	...	...	...	...	...	...	...	...	...	C..	...	...	...	...	...	...
Clone_163.13.2	...	...	...	...	...	...	...	...	...	...	...	...	...	C..	...	...	...	...	...	...
Clone_163.14.1	...	...	...	...	...	...	...	...	...	...	...	...	...	C..	...	...	...	...	...	...
Clone_163.14.2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.15.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.15.2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.16.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.AA	...	...	...
Clone_163.16.2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.AA	...	...	...
Clone_163.17.1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	..A	...	...	...
Clone_163.17.2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Clone_163.18.1	...	...	...	...	...	...	...	...	...	...	...	...	...	C..	...	...	.AA	...	...	...

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#### Appendix IV: *Triatoma* mitochondrial COI fragments of Gonzales site

Consensus_Sequence	GGT	CAA	CAA	ATC	ATA	AAG	ATA	TTG	GAA	CTC	TGT	ATT	TTC	TGT	TCG	GGG	CCT	GAG	CTG	GAA
Gonzales_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_11	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_13	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_14	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_15	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_5	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_6	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_7	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
JAM_01	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
JAM_02	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...

Consensus_Sequence	TGA	TAG	GAA	CAT	CCC	TTA	GAT	GAA	TTA	TTC	GAA	TTG	AAT	TAG	GAC	AAC	CTG	GAT	CAT	TTA
Gonzales_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	..A	.C.	A..	...	...	...
Gonzales_11	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_13	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_14	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_15	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_5	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_6	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_7	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Gonzales_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...

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[illegible]

Consensus_Sequence	TAC	TTC	TAA	CAG	ATC	GAA	ATT	TTA	ATA	CCT	CAT	TCT	TTG	ACC	CAG	CAG	GAG	GGG	GGG	ACC
JAM_02	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Consensus_Sequence	CTA	TTC	TAT	ATC	AAC	ACC	TTT	TTT	GAT	TTT	TTG	GTC	ACC	CTG	AAG	TTT				
Gonzales_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
Gonzales_11	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
Gonzales_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
Gonzales_13	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
Gonzales_14	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
Gonzales_15	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
Gonzales_2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
Gonzales_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
Gonzales_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
Gonzales_5	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
Gonzales_6	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
Gonzales_7	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
Gonzales_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
Gonzales_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
JAM_01	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				
JAM_02	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...				

# Appendix V: *Triatoma* mitochondrial COI fragments for San Marcos site 1

Consensus_sequence	GGT	CAA	CAA	ATC	ATA	AAG	ATA	TTG	GGA	CTC	TTT	ATT	TTC	TGT	TCG	GAG	CCT	GGG	CTG	GTA
MF_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_5	...	...	...	...	...	...	...	...	...	...	.A.	...	..T	...	...	...	...	...	...	...
MF_6	...	...	...	...	...	...	...	...	...	...	.A.	...	..T	...	...	...	...	...	...	...
MF_8	...	...	...	...	...	...	...	...	...	...	...	...	A..	...	..C	...	..C	...	...	...
MF_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_10	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_13	...	...	...	...	...	...	...	...	...	...	.G.	...	...	...	...	.G.	...	.A.	...	.A.

Consensus_sequence	TAA	TAG	GAA	CAT	CTC	TTA	GAT	GAA	TTA	TTC	GAA	TCG	AAT	TAG	GAC	AAC	CAG	GAT	CAT	TTA
MF_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_5	...	...	...	...	.C.	...	...	...	...	...	.T.	.T.	..C	...	.T.	...	...	...	...	...
MF_6	...	...	...	...	.C.	...	...	...	...	...	.T.	.T.	..C	...	.T.	...	...	...	...	...
MF_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.GA	...	...	...	...	...
MF_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_10	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.G.	...	...	...	...	...
MF_13	.G.	...	...	...	.C.	...	...	...	...	...	...	.T.	...	...	...	...	.T.	...	...	...

Consensus_sequence	TTG	GAG	ACG	ACC	AAA	TTT	ATA	ATG	TAG	TCG	TAA	CAG	CCC	ATG	CCT	TCG	TCA	TGA	TTT	TCT
MF_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_5	...	...	.T.	.T.	...	...	...	...	..A	...	.C.	...	.T.	...	...	.T.	.T.	.A.	...	.T.
MF_6	...	...	.T.	.T.	...	...	...	...	..A	...	.C.	...	.T.	...	...	.T.	.T.	.A.	...	.T.
MF_8	...	...	...	...	...	...	...	...	...	.T.	...	...	...	...	...	...	...	...	...	...
MF_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_10	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_13	.C.	...	...	.T.	...	...	...	...	...	.T.	...	...	...	...	.T.	...	...	.A.	...	...
Consensus_sequence	TCA	TAG	TTA	TGC	CCA	TCA	TAA	TTG	GAG	GCT	TTG	GAA	ATT	GAT	TAG	TAC	CCT	TAA	TAA	TTG
MF_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_5	.T.	...	...	.A.	.A.	.T.	.G.	...	...	.A.	...	.T.	...	...	...	.C.	..C	...	...	...
MF_6	.T.	...	...	.A.	.A.	.T.	.G.	...	...	.A.	...	.T.	...	...	...	.C.	..C	...	...	...
MF_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_10	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_12	...	...	...	...	...	.T.	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_13	...	...	...	...	.T.	.T.	...	...	...	...	...	.G.	.C.	..C	.T.	...	..C	.G.	...	...

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Consensus_sequence	TAT	ATC	CTC	CCC	TAT	CAA	GAA	ATA	TCG	CAC	ATA	GAG	GAG	CAT	CCG	TAG	ATA	TAG	CAA	TCT
MF_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_13	...	...	.C.	.TT	...	...	...	...	...	...	...	...	...	...	.T.	...	.C.	...	...	...
Consensus_sequence	TCT	CAT	TAC	ACT	TAG	CCG	GGG	TCT	CAT	CAA	TTC	TAG	GAG	CAG	TAA	ACT	TTA	TTT	CTA	CTA
MF_1	...	...	...	...	...	...	..A	...	.G.	...	...	...	...	...	...	...	...	...	...	...
MF_3	...	...	...	...	...	...	..A	...	.G.	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	..A	...	.G.	...	...	...	...	...	...	...	...	...	...	...
MF_5	...	.TC	.T.	.TC	...	.A.	.T.	.A.	.T.	...	...	...	...	.C.	...	...	.C.	.C.	.C.	.A.
MF_6	...	.TC	.T.	.TC	...	.A.	.T.	.A.	.T.	...	...	...	...	.C.	...	...	.C.	.C.	.C.	.A.
MF_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_10	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_12	...	...	...	...	...	...	.A.	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_13	.T.	...	...	..C	...	...	.T.	.T.	...	...	...	...	...	...	...	...	.C.	...	...	...
Consensus_sequence	TTA	TTA	ATA	TAC	GCC	CTG	CAG	GAA	TGC	GAC	CTG	ATC	GAA	TTC	CCT	TAT	TTG	TCT	GAT	CAG
MF_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_5	...	...	...	...	.G.	...	.T.	...	.AA	C..	.A.	...	...	.C.	.A.	...	...	.T.	...	...
MF_6	...	...	...	...	.G.	...	.T.	...	.AA	C..	.A.	...	...	.C.	.A.	...	...	.T.	...	...
MF_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_9	...	...	...	...	...	...	...	...	.A.	...	...	...	...	...	...	...	...	...	...	...
MF_10	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_13	...	...	...	...	.A.	...	...	.T.	.A.	...	.A.	...	...	...	.T.	...	...	.T.	...	.T.

Consensus_sequence	TAG	GTA	TTA	CTG	CCT	TAT	TAT	TAC	TCC	TAA	GAC	TAC	CAG	TTC	TTG	CTG	GAG	CTA	TCA	CTA
MF_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_5	.T.	.A.	...	.A.	.AC	.TC	...	...	.TT	...	.T.	.T.	.T.	...	.A.	.A.	.G.	...	.T.	.A.
MF_6	.T.	.A.	...	.A.	.AC	.TC	...	...	.TT	...	.T.	.T.	.T.	...	.A.	.A.	.G.	...	.T.	.A.
MF_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_10	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_13	.G.	.C.	...	...	.TC	...	...	...	.TT	...	...	.C.	.T.	.C.	...	.A.	...	.A.	.T.	.A.
Consensus_sequence	TGC	TCT	TAA	CAG	ATC	GAA	ACT	TTA	ATA	CTT	CAT	TCT	TTG	ATC	CAG	CCG	GAG	GGG	GGG	ACC
MF_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.A.	...
MF_5	.A.	.T.	...	...	.C.	.T.	.T.	...	...	.G.	...	...	...	.C.	.C.	.T.	.C.	...	...	...
MF_6	.A.	.T.	...	...	.C.	.T.	.T.	...	...	.G.	...	...	...	.C.	.C.	.T.	.C.	...	...	...
MF_8	...	...	...	...	.C.	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.A.	...
MF_10	.A.	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.T.
MF_13	.A.	.TC	...	...	...	...	.T.	...	...	.C.	...	...	...	.C.	...	.A.	...	...	...	...

Consensus_sequence	CAA	TTC	TTT	ACC	AAC	ACT	TAT	TCT	GAT	TTT	TTG	GTC	ACC	CTG	AAG	TTT	A
MF_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
MF_5	.T.	...	.A.	.T.	...	..C	...	...	...	...	...	...	...	...	...	...	.
MF_6	.T.	...	.A.	.T.	...	..C	...	...	...	...	...	...	...	...	...	...	.
MF_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
MF_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
MF_10	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
MF_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	.
MF_13	.T.	...	.A.	.T.	...	..C	.G.	.T.	...	...	...	...	...	...	...	...	.

## Appendix VI: *Triatoma* mitochondrial COI fragments for San Marcos site 2

Consensus_Sequence	GGT	CAA	CAA	ATC	ATA	AAG	ATA	TTG	GGA	CTC	TGT	ATT	TTC	TGT	TCN	GGG	GCC	TGA	GCT	GGA
Trial_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	..G	...	...	...	...	...
Trial_2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	..-	...	...	...	...	...
Consensus_Sequence	ATG	ATA	GGA	ACA	TCC	CTT	AGA	TGA	ATT	ATT	CGA	ATT	GAA	TTA	GGA	CAA	CCT	GGA	TCA	TTT
Trial_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Trial_2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Consensus_Sequence	ATC	GGA	GAC	GAT	CAA	ATT	TAT	AAT	GTA	GTT	GTA	ACA	GCC	CAT	GCT	TTC	GTC	ATA	ATT	TTC
Trial_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Trial_2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Consensus_Sequence	TTC	ATA	GTT	ATG	CCT	ATT	ATA	ATT	GGA	GGC	TTT	GGG	AAC	TGA	CTT	GTA	CCC	CTA	ATA	ATT
Trial_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Trial_2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Consensus_Sequence	GGT	GCC	CCA	GAT	ATA	GCT	TTC	CCT	CGA	ATA	AAT	AAT	ATA	AGA	TTT	TGA	CTC	TTA	CCC	CCA
Trial_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Trial_2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Consensus_Sequence	GCC	CTC	ACC	CTT	TTA	TTA	GTA	AGA	AGA	CTT	GTA	GAA	AGA	GGG	GCA	GGA	ACA	GGA	TGA	ACA
Trial_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Trial_2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Consensus_Sequence	GTA	TAT	CCC	CCT	TTA	TCA	AGA	AAT	ATC	GCA	CAT	AGA	GGA	GCA	TCT	GTA	GAC	ATA	GCA	ATC
Trial_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Trial_2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Consensus_Sequence	TTT	TCA	TTA	CAC	CTA	GCT	GGT	GTT	TCA	TCA	ATT	CTA	GGA	GCA	GTA	AAC	TTC	ATT	TCT	ACT
Trial_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
Trial_2	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...

Consensus_Sequence	ATT ATT AAT ATA CGA CCT GCA GGT ATA CGA CCA GAT CGA ATT CCT TTA TTT GTT TGA TCT
Trial_1	... ..
Trial_2	... ..
Consensus_Sequence	GTG GGC ATT ACT GCT CTA TTA TTA CTT TTA AGA CTC CCT GTC CTT GCA GGA GCA ATT ACA
Trial_1	... ..
Trial_2	... ..
Consensus_Sequence	ATA CTT CTA ACA GAT CGA AAT TTT AAT ACC TCA TTC TTT GAC CCA GCA GGA GGG GGG GAC
Trial_1	... ..
Trial_2	... ..
Consensus_Sequence	CCT ATT CTA TAT CAA CAC CTG TTT TGA TTT TTT GGT CAC CCT GAA GTT TA
Trial_1	... ..
Trial_2	... ..

# Appendix VII: *Triatoma* Cytochrome B fragments for San Marcos site 1

MF_1	GGA	CGW	GGW	ATT	TAT	TAT	GGA	TCC	TAT	AAG	CTC	TTT	ATA	ACC	TGA	GCA	GTA	GGT	GTT	ATT
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_5	...	...	...	...	...	...	...	..A	...	..A	..T	C.G	...	...	...	AT.	A.C	..A	..A	C.A
MF_6	...	...	...	...	...	...	...	..A	...	..A	..T	C.G	...	...	...	AT.	A.C	..A	..A	C.A
MF_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	A..	...	...	...
MF_10	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	..G	...	...	...	...
MF_13	...	...	...	...	...	...	...	...	..C	..A	...	..C	...	...	...	A..	...	..G	...	...
MF_1	ATT	TTA	TTT	ATC	ACT	ATA	GGA	GCC	GCA	TTC	CTA	GGA	TAT	GTT	CTT	CCC	TGA	GGG	CAA	ATA
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_5	C..	...	...	..T	..T.	...	..G	A..	..C	...	..G	...	...	..C	..C	..A	...	..A	...	...
MF_6	C..	...	...	..T	..T.	...	..G	A..	..C	...	..G	...	...	..C	..C	..A	...	..A	...	...
MF_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	..G	...	...	..G
MF_9	...	...	...	...	...	...	..G	...	...	...	...	...	...	...	...	...	...	...	..G	...
MF_10	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	..A	...	..G	...
MF_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	..A	...	..G	...
MF_13	...	C.C	...	..A	...	...	...	..T	...	...	..G	...	...	...	..C	...	...	..A	...	...
MF_1	TCT	TTA	TGG	GGG	GCC	ACA	GTT	ATT	ACT	AAT	TTA	ATA	TCC	GCC	ATC	CCT	TAC	CTA	GGA	AAC
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_5	..A	C.C	..A	..A	..A	...	..A	...	...	...	C.C	T..	..A	..T	...	...	..T	T..	...	...
MF_6	..A	C.C	..A	..A	..A	...	..A	...	...	...	C.C	T..	..A	..T	...	...	..T	T..	...	...
MF_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_10	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	..T
MF_12	...	...	...	..A	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_13	..C	..G	..A	..A	...	...	...	...	...	..C	...	...	...	...	...	...	A.T	T.G	...	...

MF_1	GAT	TTA	GTT	AAA	TGA	TTA	TGA	GGG	GGA	TTC	TCT	GTT	GAT	AAC	GCC	ACC	TTG	ACT	CGA	TTT
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_5	..A	C..	..C	...	...	...	...	...	...	..T	..C	...	..C	..T	..T	...	C.C	...	...	...
MF_6	..A	C..	..C	...	...	...	...	...	...	..T	..C	...	..C	..T	..T	...	C.C	...	...	...
MF_8	...	...	...	...	...	...	...	...	...	...	...	..C	...	...	...	...	...	...	...	...
MF_9	...	...	...	...	...	...	...	...	...	...	...	..C	...	...	...	...	...	...	...	...
MF_10	...	...	...	...	...	...	...	...	..G	...	...	..C	...	...	...	...	...	...	...	...
MF_12	...	...	...	...	...	...	...	...	..G	...	...	..A	...	...	...	...	...	...	...	...
MF_13	...	C..	...	...	...	...	..G	..A	...	..T	..C	..A	...	...	..T	...	..A	...	..C	...

	TTC	GCC	CTC	CAC	TTT	CTA	CTA	CCC	TTT	ATT	ATT	GCA	GCT	ATG	GTA	ATA	ATC	CAT	CTT	TTA
MF_1																				
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_5	..T	...	..A	..T	..C	..T	...	...	...	...	...	..T	..A	C.A	..T	...	G.A	..C	..A	C..
MF_6	..T	...	..A	..T	..C	..T	...	...	...	...	...	..T	..A	C.A	..T	...	G.A	..C	..A	C..
MF_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_10	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_13	..T	...	..T	...	...	..C	T..	...	...	...	...	...	..C	..A	...	...	...	..C	...	...

[illegible]

MF_1	TTT	TTA	CAC	CAA	ACA	GGA	TCT	AGA	AAC	CCA	TTA	GGG	TTG	AAT	AGA	AAC	TTT	GAT	AAA	ATT
MF_13	..C	...	..T	...	...	...	..A	...	...	...	...	..A	C.A	...	...	...	...	...	...	...

	CCA	TTT	CAC	CCT	TAT	TTC	TCC	ATC	AAG	GAT	TTA	ATA	GGA	GTA	TCA	CTA	ACA	TTA	ATA	TTT
MF_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_5	...	...	...	..A	..C	...	..T	...	..A	..C	...	...	...	..C	AT.	T..	..C	C.T	...	...
MF_6	...	...	...	..A	..C	...	..T	...	..A	..C	...	...	...	..C	AT.	T..	..C	C.T	...	...
MF_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_10	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_13	...	..C	..T	..A	..C	...	...	...	..A	..C	C..	...	...	...	..C	T..	..C	...	...	...

	TTT	ATT	CTA	TTA	AGC	TTA	TGA	GAG	GCC	CCA	ATT	CTG	ATA	GAC	CCA	GAA	AAT	TTT	ATT	CCT
MF_1	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_5	...	...	.C	C.C	.AT	...	.G	.A	C.A	.G.	...	...	GG.	.T	...	...	.C	.C	.C	.A
MF_6	...	...	.C	C.C	.AT	...	.G	.A	C.A	.G.	...	...	GG.	.T	...	...	.C	.C	.C	.A
MF_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_10	...	...	.G	...	...	...	...	...	...	...	...	.A	...	...	...	...	...	...	...	...
MF_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_13	.C	.C	.C	...	.T	C.T	.G	.A	...	...	.C	.A	...	.T	...	...	.C	.C	...	.A

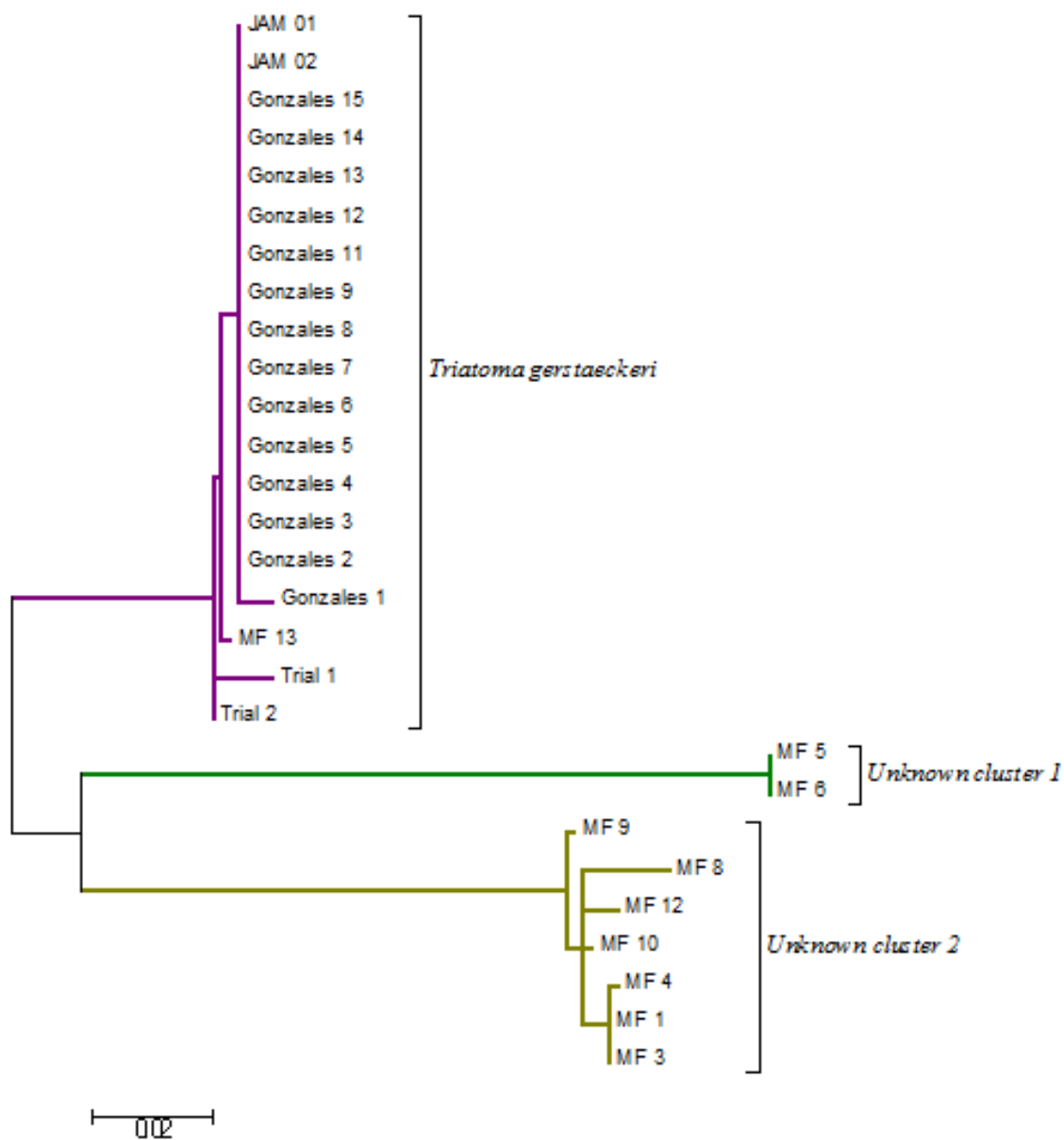
[illegible]

MF_1	GCA AAC CCA TTG GTA ACA CCA GTG CAC ATT CAA CCA GAA TGA TAC TTC CTA TTT GCA TAC
MF_13	... ..T ... ..A ..T ... ..T ... ..T ... ..T ... ..T ... ..C ... ..
MF_1	GCA ATT TTA CGA TCC ATT CCT AAT AAG TTA GGA GGG GTC ATT GCA ATA GTC TCA TCA ATC
MF_3	... ..
MF_4	... ..
MF_5	..C ... ..C ... ..C ..A ... ..T ..A ... ..T G.. ..C ..T
MF_6	..C ... ..C ... ..C ..A ... ..T ..A ... ..T G.. ..C ..T
MF_8	... ..
MF_9	... ..
MF_10	... ..
MF_12	... ..
MF_13	... ..C ... ..A C.. ..A ..T ... ..A ..T ... ..A ... ..T
MF_1	GCA ATT ATT TTA ATC CTT CCA TTC ACT AAC AAA AGA AAA TTT CAA GGC CTC CCA TTT TAC
MF_3	... ..
MF_4	... ..
MF_5	... ..C ... C.T ..T ..A ... ..T ... ..A ..A T.. ...
MF_6	... ..C ... C.T ..T ..A ... ..T ... ..A ..A T.. ...
MF_8	... ..
MF_9	... ..
MF_10	... ..
MF_12	... ..
MF_13	..C ... ..C ... GAT ... ..T ... ..T ... ..C ... ..A.C ..C ...

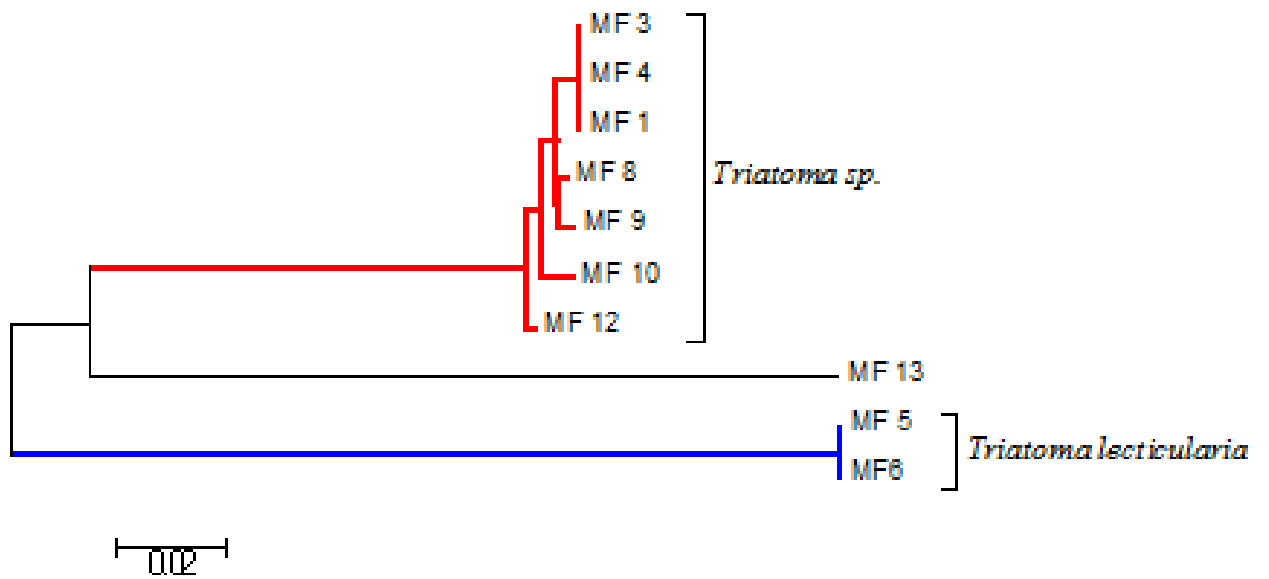
MF_1	CCA	ATT	AGA	CAA	GTT	ATA	TTT	TGA	GCA	CTC	GCA	GTT	ATT	TTA	ATC	TTA	CTA	ACC	TGA	ATT
MF_3	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_4	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_5	...	..A	.AT	...	A..	...	..C	...	T.C	..A	A.T	.C.	...	C..	..T	...	...	...	...	...
MF_6	...	..A	.AT	...	A..	...	..C	...	T.C	..A	A.T	.C.	...	C..	..T	...	...	...	...	...
MF_8	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_9	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_10	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	Y..	..Y	...	...
MF_12	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
MF_13	...	T.A	GAC	A..	T.A	TAT	.C.	GA.	CAC	T.A	CA.	C.A	T.C	.AT	T.T	ACT	AAC	CTG	AAT	TGG

MF_1	GGA	GC
MF_3	...	..
MF_4	...	..
MF_5	...	..
MF_6	...	..
MF_8	...	..
MF_9	...	..
MF_10	..W	..
MF_12	...	..
MF_13	A.C	..

**Appendix VIII:** Phylogenetic map of *Triatoma* samples using mitochondrial COI



**Appendix IX:** Phylogenetic tree of *Triatoma* samples using Cytochrome B



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